

Multi-drug resistant of *Escherichia coli* isolated from urinary tract infections

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Abstract

Urinary tract infection (UTI) is one of the commonest bacterial infections caused by microbial invasion of tissue lining the urinary tract. *Escherichia coli* (*E. coli*) is the primary etiologic agent of UTI, also antimicrobial resistance is an evolving and growing problem in UTI. The aim of this study is to determine the prevalence of antibiotic resistant of *E. coli* among outpatients with UTI in Mukalla city, Hadhramout-Yemen. Mid-stream urine specimens were collected, aseptically cultured, and the isolates were identified by using standard microbiological techniques. Antimicrobial susceptibility test was performed by disk diffusion method. Of the 295 urine samples, the total growth *E. coli* was 29 (78.38%) of the total positive samples (37). Of total antibiotics used, amoxicillin/clavulanic acid, cefotaxime and cefuroxime sodium showed the highest resistance to *E. coli* with 82.76%, 72.41% and 65.52% respectively, while 75.86% of the isolates were susceptible to co-trimoxazole. The study revealed that high resistant and multi-resistant of the urinary *E. coli* isolates to antibiotics. Co-trimoxazole was the most effective antibiotic on *E. coli* isolates in this study.

Key words: UTI, *E. coli*, Antimicrobial susceptibility, Outpatients.

Introduction

Urinary tract infection (UTI) is one of the commonest domiciliary and nosocomial bacterial infections caused by microbial invasion of tissue lining the urinary tract. It refers to the presence of significant bacteruria and pyuria in the midstream sample of urine¹⁸. *Escherichia coli* (*E. coli*) being the primary etiologic agent, causing UTI among both inpatients and outpatients^{10,11}, is also accounted for approximately 90% of first UTI in young women, the symptoms and signs include urinary frequency, dysuria, hematuria, and pyuria; none of these symptoms and signs are specific for *E. coli* infections⁸. Antimicrobial resistance is an evolving and growing problem in UTI. Of more concern is the increasing incidence of infections caused by strains of *E. coli* that are resistant to commonly used antimicrobial agents, especially to trimethoprim, sulphamethoxazole and beta-lactam antibiotics²¹. The extensive and inappropriate use of antimicrobial agents has invariably resulted in the development of bacterial resistance which has become a major problem, therefore it is necessary for continuous surveillance of antimicrobial resistance among these organisms. The aim of this study is to determine the prevalence of *E. coli* and to assess the level of drug resistance in outpatients with UTI in Mukalla city, Hadhramout - Yemen.

Materials and methods

A total of 295 of midstream urine were obtained from outpatients in Mukalla city in a period from January to July 2014. The samples were collected into sterile plastic disposable bottles, then inoculated by calibrated loop capable of delivering 0.001 ml of urine on MacConkey agar (Oxoid) which were incubated aerobically at 37°C for 24 hours. The number and types of colonies grown on the medium was recorded as being insignificant when samples gave a colony count of less than 10⁴ CFU ml, while samples with colony count equal to or greater than approximately 10⁵ CFU ml of the urine samples were considered to have significant bacteruria⁶. Bacterial isolates were identified by conventional diagnostic methods, and antibiotic susceptibility test carried out using disc diffusion method on Mueller-Hinton agar (Oxoid) that has been performed by Clinical

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Laboratory Standards Institute (CLSI). The antibiotic discs used were cefuroxime sodium (30µg), cefotaxime (30µg), amoxicillin/clavulanic acid (30µg), ciprofloxacin (5µg), amikacin (30µg), lincomycin (15µg) and co-trimoxazole (25µg). A suspension of tested organism was adjusted against 0.5 MacFarland standard turbidity and inoculated into media, then incubated at 35-37 °C for 16-18 hours and examined for evidence of growth. Interpretation as 'sensitive', 'moderate sensitive' or 'resistant' was done on the basis of the diameters of zones of inhibition of bacterial growth as recommended by the disc manufacturer (Oxoid).

Results and discussion

Of the 295 samples of urine examined, 37(12.54%) showed significant bacteruria; 26 (8.81%) had insignificant bacteruria, while 232 (78.65%) had no growth (Table 1). *E. coli* was the most common bacterial isolated from the tested individuals with a frequency of occurrence at 29 (78.38%), Table (2). Similar findings regarding the frequency of urinary *E. coli* infection have been observed by other researchers carried out in the European countries and North American¹⁹, Lahore¹⁷, Saudi Arabia²³ and India²⁹. Another study reported that the most prevalent organism isolated from urine was *E. coli* 86.02% in Nepal³¹, 52% in Tikrit, Iraq¹² and 73.0% in Poland¹⁵. The results of antimicrobial sensitivity test revealed that the most resistance rates of *E. coli* are as follows: 82.76% to amoxicillin/clavulanic acid, 72.41% to cefotaxime, 65.52% to cefuroxime sodium, 41.38% to amikacin and lincomycin, (Table 3). This pattern of resistance is comparable to a study carried out locally in which *E. coli* showed a high resistance against quinolones with 84.6% and penicillin 78.8%² as well as other studies carried out abroad such as in Bosnia²⁰, Saudi Arabia²², Palestine⁹ and Nigeria⁷.

Regarding to multi-resistant of antimicrobial agents, the organism is considered as multi-resistant if it is resistant to three or more antimicrobials³⁰. In this study, susceptibility test showed that the multi-resistance rate among the isolates of *E. coli* was observed (table 3). These results approximately agreed with those studies that showed a significant of high level multi-resistance of antibiotics to *E. coli* in several studies in Spain 20.6%²⁶, the USA 7.1%²⁷ and Jordan 59.9%³. We suggest the high level of resistance of *E. coli* due to easy purchase antibiotics without prescription, not restricted with time and dose of antibiotic, culture and sensitivity test neglected and the empirical treatment dependently.

From this study, it can be seen that co-trimoxazole, followed by ciprofloxacin, were virtually used against urinary *E. coli* as they were effective against 75.86% and 37.93% of all the isolates respectively, while amikacin and lincomycin were slightly better and showed moderate activity against 51.72% of isolates (Table 3). These results were approximately agreed with those in a study that showed the percentages of sensitivity of urinary *E. coli* isolates to co-trimoxazole and ciprofloxacin were 62.1% and 47.1% respectively⁵, other studies showed that the overall susceptibilities of *E. coli* was 58% for ciprofloxacin²⁵. Other study showed decreased the percentages of sensitivity of *E. coli* to ciprofloxacin 43% and to co-trimoxazole 13%²⁸. A study carried out in Pakistan it was observed that 16.6% of total tested urinary *E. coli* isolates were sensitive to ciprofloxacin and 13.95% to co-trimoxazole¹³, while other studies showed increase of the percentages of sensitivity of *E. coli* to ciprofloxacin and co-trimoxazole, such as in Spain, the sensitivity to co-trimoxazole and ciprofloxacin was found at rates of 67.4% and 80.7% respectively²⁶. A study carried out in Hong Kong revealed that ciprofloxacin sensitivity was 77.9% and co-trimoxazole was 66% of *E. coli* isolates¹⁴. Other studies documented that 63% sensitivity of *E. coli* isolates to ciprofloxacin²⁴. A study carried out in India showed that more than 70% were sensitive to ciprofloxacin²⁹. Other studies revealed that high sensitivity rates of *E. coli* strains were observed to ciprofloxacin (85.5%) in Jordan⁴. In another study, ciprofloxacin at the rate of 90.4% was considered as the most active agent against *E. coli* isolates¹.

In our results, amikacin showed moderate sensitivity against *E. coli* isolates (51.72%), other studies documented that sensitivity of *E. coli* isolates to amikacin was 88%²⁴. A study carried out in India showed that more than 80% of the *E. coli* isolates were sensitive to amikacin²⁹. Where as other studies revealed that high sensitivity rates for amikacin 100% were observed in Jordan⁴. In

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 another study, amikacin was the most active agent against *E. coli* isolates (82.9%)¹. Also, high sensitivity of *E. coli* to amikacin was observed in Baghdad Hospital, Iraq (94%)¹⁶.

Conclusion

This study revealed high level resistance of the urinary *E. coli* isolates as the problem of resistance and multi-resistance in outpatients UTI. According to our results, co-trimoxazole is the most effective antibiotic against *E. coli* isolates.

Table 1. Number of urine samples examined during study period

	No.	%
Positive samples (with significant bacteruria)	37	12.54
Negative samples (samples non-significant bacteruria)	26	8.81
Number of samples with no growth	232	78.65
Total	295	100.00

Table 2. *Escherichia coli* isolated from urine samples

Isolates	No.	%
<i>Escherichia coli</i>	29	78.38
Others	8	21.62
Total	37	100.0

Table 3. Antibiotic susceptibility patterns of *Escherichia coli* isolates

Antibiotic	Sensitive (%)	Moderate (%)	Resistant (%)
Cefuroxime sodium	3(10.34)	7(24.14)	19(65.52)
Cefotaxime	2(6.90)	6(20.69)	21(72.41)
Amoxicillin/clavulanic acid	2(6.90)	3(10.34)	24(82.76)
Ciprofloxacin	11(37.93)	11(37.93)	7(24.14)
Amikacin	2(6.90)	15(51.72)	12(41.38)
Lincomycin	2(6.90)	15(51.72)	12(41.38)
Co-trimoxazole	22(75.86)	3(10.34)	4(13.80)

References

1. Afsharpaiman S., Bairaghdar F., Torkaman M., Kavehmanesh Z., Amirsalari S., Moradi M., and Safavimirmahalleh M.J. (2012). Bacterial Pathogens and Resistance Patterns in Children With Community-Acquired Urinary Tract Infection: A Cross Sectional Study. *J. Compr. Ped.* Vol. 3(1); 16-20.
2. Al-Haddad A.M., BinGhouth A.S. and El-Hosseiny M. (2010). Microbial Resistance In Patients With Urinary Tract Infections In Al-Mukalla, Yemen. *Sudan J.M.S.* Vol. 5,(2); 145-150.
3. Al Mardeni R.I., Batarseh A., Omaish L., Shraideh M., Batarseh B. and Unis N. (2009). Empirical treatment for pediatric urinary tract infection and resistance patterns of uropathogens in Queen Alia hospital and prince A'isha military Center-Jordan. *Saudi J. Kidney Dis. Transpl.* Vol. 20(1); 135-139.
4. Alshara M. (2011). Antimicrobial Resistant Pattern of *Escherichia coli* Strains Isolated from Pediatric Patients in Jordan. *ActaMedicalIranica.* Vol. 49(5); 293-295.
5. Ashraf V. and Satti L. (2014). In-vitro efficacy of nitrofurantoin, ciprofloxacin and co-trimoxazole against various urinary isolates. *Gomal Journal of Medical Sciences.* Vol. 12(4); 201-204.

6. Ayoade F., Moro D.D. and Ebene O.L. (2013). Prevalence and Antimicrobial Susceptibility Pattern of Asymptomatic Urinary Tract Infections of Bacterial and Parasitic Origins among University Students in Redemption Camp, Ogun State, Nigeria. *Open Journal of Medical Microbiology*. Vol. 3; 219-226.
7. Bakare R.A., Oni A.A., Arowojolu A.O., Ayuba T.T. and Toki R.A. (1999). *In vitro* activity of Pefloxacin and other antibiotics against Gram-negative bacteria in Ibadan, Nigeria. *Nigerian Quarterly J. Hospital Med.* Vol. 9(1); 54-57.
8. Brook F.G., Carroll K.C., Butel J.S., Morse S.A. and Mietzner T.A. (2010). Enteric Gram-negative rods (Enterobacteriaceae). In Jawetz, Melnick and Adelberg's *Medical Microbiology*. New York: MacGraw-Hill. 213-226.
9. El-Astal Z. (2004). Bacterial pathogens and their antimicrobial susceptibility in Gaza strip, Palestine. *Pakistan J. Med.* Vol. 20(4); 365-370.
10. Fadda G., Nicoletti G., Schito G.C. and Tempera G. (2005). Antimicrobial susceptibility patterns of contemporary pathogens from uncomplicated urinary tract infections isolated in a multicenter Italian survey: possible impact on guidelines. *J. Chemother.* Vol. 17(3); 251-57.
11. Farrel D.J., Morrissey I., Rubies D., Robbins M. and Felmingham D.A. (2003). UK multi-center study of the antimicrobial susceptibility of bacterial pathogens causing UTI. *J. Infect.* Vol. 46; 94-100.
12. Hamada T.A., Mahmood A.R. and Ahmed I.A. (2008). Antibiotic Resistance In Pathogenic Bacteria Isolated From UTIs In Tikrit Province. *Tikrit Medical Journal*. Vol. 14(1); 203-210.
13. Hameed S., Afzal T., Cheema S.M., Momina A., and Hussain A. (2012). Distribution and susceptibility pattern of urinary tract bacterial pathogens in an outpatients setting. A laboratory based study–Faisalabad. *JUMDC*. Vol. 3(2); 27-34.
14. Hoa P.L., Wong R.C., Yip K.S., Loke S.L., Leung M.S., Mak G.C., Chow F.K., Tsang K.W., Que T.L, and Combat study group. (2007). Antimicrobial resistance in *Escherichia coli* outpatient urinary isolates from women: emerging multidrug resistance phenotypes. *Diagn. Microbiol. Infect. Dis.* Vol. 59(4); 439-445.
15. Hryniewicz K., Szczypka K., Sulikowska A., Jankowski K., Betlejewska K. and Hryniewicz W. (2001). Antibiotic Susceptibility Of Bacterial Strains Isolated From Urinary Tract Infections In Poland. *Journal of Antimicrobial Chemotherapy*. Vol. 47; 773–780.
16. Hussein N.S. (2014). Clinical, Etiology and Antibiotic Susceptibility Profiles of Community-Acquired Urinary Tract Infection in a Baghdad Hospital. *Med. Surg. Urol.* Vol. 3(2); 136.
17. Javeed I., Hafeez R. and Anwar M.S. (2011). Antibiotic Susceptibility Pattern Of Bacterial Isolates From Patients Admitted To A tertiary Care Hospital In Lahore. *Biomedica*. Vol. 27; 19-23.
18. Jha N. and Bapat S.K. (2005). A study of sensitivity and resistance of pathogenic microorganisms causing UTI in Kathmandu valley. *KUMJ*. Vol. 3; 123-129.
19. Jones ME., Draghi DC., Thornsberry C., Karlowsky J.A., Sahm D.F. and Wenzel R.P. (2004). Emerging Resistance Among Bacterial Pathogens In The Intensive Care Unit European And North American Surveillance Study (2000-2002). *Ann. Clin. Microbiol. Antimicrob.* Vol. (3); 14.
20. Kamberovic S.U. (2006). Antibiotic resistance of coliform organisms from community – acquired urinary tract infections in Zenica-Doboj Canton, Bosnia and Herzegovina. *J. Antimicrobial Chemotherapy*. Vol. 58; 344-348.
21. Kahlmeter G. (2003). An international survey of the antimicrobial susceptibility of pathogens from uncomplicated UTIs: the ECO.SENS Project. *J. Antimicrob. Chemother.* Vol. 51(1); 69-76.
22. Khameneh Z.R., and Afshar A.T. (2009). Antimicrobial susceptibility patterns of urinary tract pathogens. *Saudi J. Kidney Dis. Transpl.* Vol. 20; 251-253.
23. Masoud E.A., Mahdy M.E. and Esmat A.M. (2011). Bacterial Prevalence and Resistance to Antimicrobial Agents in Southwest, Saudi Arabia. *Egypt. Acad. J. biolog. Sci.* Vol. 3(1): 105-111.

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24. Naeem M., Khan M.A. and Qazi S.M. (2010). Antibiotic Susceptibility Pattern of Bacterial Pathogens Causing Urinary Tract Infection in a Tertiary Care Hospital. Ann. Pak. Inst. Med. Sci. Vol. 6(4); 214-218.
25. Ohieku J.D. and Magaji R.A.(2013).Urinary Tract Infections Associated with *Escherichia Coli*: A 2005 to 2009 Clinical Assessment of Trends in Fluoroquinolones Activities in Maiduguri-City, Nigeria. Journal of Applied Pharmaceutical Science. Vol. 3(08); 084-091.
26. Oteo J., Lazaro E., de Abajo F.J., Baquero F., Campos J. and Spanish members of EARSS. (2005). Antimicrobial-resistant Invasive *Escherichia coli*, Spain. Emerg. Infect. Dis. Vol. 11(4); 546–553.
27. Sahm D.F., Thorensberry C., Mayfield D.C., Jones M.E. and Karlowsky J.A. (2001). Multidrug-resistance urinary tract isolates of *Escherichia coli*: Prevalence and patient demographics in the United States in 2000. Antimicrob. Agents Chemother. Vol. 45(5); 1402-1406.
28. Shaikh D., Ashfaq S., Shaikh K., Shaikh M., Naqvi B.S., Mahmood Z.A. and Majid R. (2005). Studies on Resistance/Sensitivity Pattern of Bacteria Related With Urinary Tract Infections. Medical Journal Of Islamic World Academy of Sciences. Vol. 15(4); 129-133.
29. Shalini, Joshi MC, Rashid MK and Joshi HS. (2011). Study of Antibiotic Sensitivity Pattern In Urinary Tract Infection At A Tertiary Hospital. N.J.I.R.M. Vol. 2(3); 43-46.
30. Taneja N., Mohan B., Khurana S. and Sharma M. (2004). Antimicrobial resistance in selected bacterial enteropathogens in north India. Indian J. Med. Res. Vol. 120; 39-43.
31. Thakur P., Ghimire P., Rijal K.R., and Singh G.K. (2012). Antimicrobial Resistance Pattern Of *Escherichia coli* Isolated From Urine Samples In Patients Visiting Tertiary Health Care Centre In Eastern Nepal. Sunsari Technical College Journal. Vol. 1(1); 22-26.

المقاومة المتعددة لجراثيم الايشريكية القولونية المعزولة من عدوى المسالك البولية

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الملخص

يُعد التهاب المسالك البولية أحد العدوى الجرثومية الشائعة المتسببة باختراق الميكروبات للطبقات المبطنة للجهاز البولي. وتُعد جراثيم الايشريكية القولونية المسبب الأساسي لالتهابات المسالك البولية وبعد ظهور سلالات لهذه الجراثيم مقاومة للمضادات الحيوية مشكلة متنامية في التهابات المسالك البولية. هدفت هذه الدراسة لتحديد انتشار العدوى بجراثيم الايشريكية القولونية وحساسيتها للمضادات الحيوية في مرضى العيادات الخارجية في مدينة المكلا ، حضرموت – اليمن. جمعت عينات البول من منتصف السبيل بطريقة معقمة ثم زرعت على الأوساط الزراعية وتم التعرف على العزلات الجرثومية بتقنيات التشخيص الجرثومية في المختبر وتم إجراء فحص الحساسية للمضادات الحيوية بطريقة الانتشار من الأقراص. بلغ عدد جراثيم الايشريكية القولونية المعزولة 29 (78,38%) من إجمالي 295 عينة، ومن إجمالي المضادات الحيوية المستخدمة في الدراسة تبين أن جراثيم الايشريكية القولونية كانت مقاومة للمضادات الحيوية أموكسيسيلين/حمض الكلافولينك وسيفوتاكسيم وسيفيوروكسيم الصوديوم 38,33%، 77,78%، 66,67% على التوالي، في حين بلغت حساسية العزلات الجرثومية للمضاد الحيوي كوترايموكسازول 77,78%. يستنتج من هذه الدراسة وجود مقاومة عالية ومقاومة متعددة للمضادات الحيوية بشكل شائع لعزلات الايشريكية القولونية البولية، وأن المضاد الحيوي كوترايموكسازول هو الأكثر فعالية على هذه العزلات في هذه الدراسة.

الكلمات المفتاحية: التهابات المسالك البولية، جراثيم الايشريكية القولونية، حساسية المضادات الحيوية، مرضى العيادات الخارجية.